Efficacy of treatment and survival rate of foals with pneumonia: Retrospective comparison of rifampin/ azithromycin and rifampin/tulathromycin

Denise Arnold-Lehna¹, Monica Venner², Londa J. Berghaus³, Roy Berghaus⁴ and Steeve Giguère³

¹ Equine Clinic, School of Veterinary Medicine, Hannover, Germany

² Veterinary Clinic, Destedt, Germany

³ Large Animal Medicine, University of Georgia, Athens, GA, USA

⁴ Department of Population Health, University of Georgia, Athens, GA, USA

Summary: Rhodococcus equi (R. equi), a Gram-positive facultative intracellular pathogen is one of the most common causes of pneumonia in foals. The retrospective study compares the efficacy of rifampin/azithromycin versus rifampin/tulathromycin for the treatment of R. equi pneumonia in foals. Weekly medical data of foals born in the seasons 2012 to 2016 (i.e. five foaling seasons) that developed pneumonia were collected and analysed. Foals 21 days of age or older with a pulmonary consolidation of more than 1 cm (abscess score \geq 1 cm) were enrolled for participation in the study. All foals meeting the criteria for inclusion were listed in a spreadsheet and 330 foals affected with pneumonia each year were randomly selected by an online research randomizer. Foals with incorrect or missing data and those that were included in previous treatment studies were excluded from the analysis, resulting in a total enrolment of 1544 foals with pneumonia. The farm established a screening program in 2003 in order to detect foals in the early course of pneumonia, so all foals were examined once weekly starting at the age of two weeks until four and a half months of age. A physical examination of the respiratory tract, body temperature, haematology and an ultrasonographic examination of the lungs were included. Sonography areas with visible consolidation were measured and added to calculate an "abscess score" which represents the extent of pulmonary damage. Weekly medical data were analysed retrospectively. The risk of therapy failure, which means either death or worsening of pneumonia requiring that the foal be switched to another therapy, showed in a univariate comparison of the two treatments, that there was a higher incidence of treatment failure in foals treated with RIF/AZM (32/353, 9.1%) than in foals that were treated with RIF/TUL (19/406, 4.7%; Pearson's χ^2 , P = 0.016). In a Kaplan-Meier analysis, there was also a significant difference between the two treatments with respect to their survival functions (Log-rank test, P = 0.045; Figure 1). The incidence of mortality due to R. equi pneumonia was higher in foals that were treated with RIF/AZM (8/353, 2.3%) than in those that were treated with RIF/TUL (0/406, 0.0%; Fisher's exact test, P = 0.002). After adjusting for year, age of foals at the beginning of treatment, and abscess score at the beginning of treatment, the risk of treatment failure was not significantly lower for foals that were treated with RIF/TUL compared to those that were treated with RIF/AZM [Hazard Ratio (95% CI) = 0.75 (0.38, 1.5)]. Age of foals was negatively associated with treatment failure, with the risk of failure decreasing by 30% for every one-month increase in age at the beginning of treatment [HR (95% CI) = 0.70 (0.54, 0.91)]. Abscess score was positively associated with treatment failure, with the risk of failure increasing by 4% for every one cm increase in score [HR (95% CI) = 1.04 (1.01, 1.07)]. Year was also associated with the risk of treatment failure but it did not meet the proportional hazards assumption, so the model was adjusted for the effects of year by stratification. In conclusion Rifampin/tulathromycin is an effective therapy against mild to moderate R. equi pneumonia in foals as this treatment has similar survival rates compared to rifampin/azithromycin. In terms of costs and dosage form, the combination of tulathromycin with rifampin is an adequate therapeutic option for the treatment of foals with R. equi pneumonia.

Keywords: antibiotics, foal, pneumonia, Rhodococcus equi

Citation: Arnold-Lehna D., Venner M., Berghaus L. J., Berghaus R., Giguére S. (2019) Efficacy of treatment and survival rate of foals with pneumonia: Retrospective comparison of rifampin/azithromycin and rifampin/tulathromycin. Pferdeheilkunde 35, 423–430; DOI 10.21836/ PEM20190504

Correspondence: PD Dr. Monica Venner, Equine Clinic, Trift 4, 38162 Destedt, Germany; mvenner@gmx.de

Received: June 20, 2019 | Accepted: July 17, 2019

Introduction

Rhodococcus equi (R. equi), a Gram-positive facultative intracellular pathogen is one of the most common causes of pneumonia in foals, in particular those between one and six months of age. Although R. equi can be cultured from the environment of many stables, the clinical disease in foals is endemic at some farms, sporadic at others, and unrecognized at many (Cohen 2014). On farms where the disease is endemic, costs associated with morbidity and mortality attributable to R. equi may be very high. The treatment of abscessing pneumonia in foals is challenging due to the emergence of resistant strains of the bacterium against standard antimicrobial drugs and the lack of alternatives for efficient treatment or prevention (e.g. vaccination) (Peters et al. 2011, Berlin et al. 2016). Foals acquire R. equi infection by inhalation of dust and soil particles carrying virulent bacteria (Vázguez-Boland 2010). Compared to extracellular pathogens as Streptococcus equi subsp. zooepidemicus (S. zooepidemicus), the survival strategy of R. equi is based on the intracellular survival and reproduction especially within alveolar macrophages after phagocytosis (Giguère et al. 2012, Muscatello 2012, Giguère et al. 2015). Macrolide antibiotics are a major part of the standard therapy (Muscatello 2012), as they widely distribute into pulmonary compartments (Gaynor and Mankin 2003). In foals, macrolides show unique pharmacokinetic features such as rapid and extensive distribution and long persistence in pulmonary epithelial lining fluid (PELF) and in bronchoalveolar cells (Villarino et al. 2013). For all the macrolides, the rate of elimination is slow, and drugs tend to persist for a long period of time in bronchoalveolar cells when compared with PELF and plasma (Villarino et al. 2013).

The combination of macrolides with rifampin achieves a synergism in efficacy against R. equi in vitro as well as in vivo (Prescott and Nicholson 1984, Nordmann and Ronco 1992, Giguère et al. 2012). In addition, the use of the two classes of drugs in combination reduces the likelihood of development of resistance to either drug (Prescott and Nicholson 1984, Nordmann and Ronco 1992, Berghaus et al. 2013). It is known that the development of antimicrobial resistance to rifampin is relatively rapid when used in monotherapy treatment regimens (Giguère et al. 2010). This is particularly important given the fact that the prevalence of macrolide resistance among R. equi isolates is increasing (Giguère et al. 2010). Some studies describe the emergence of widespread macrolide and rifampin resistance at a farm as a consequence of wide-spread use of these drugs in subclinically affected animals (Hoffman et al 1993, Burton et al. 2013). The best method to prevent or minimize the development of antimicrobial resistance is to limit the use of antimicrobial agents to only those individuals that require treatment. (Weese et al. 2015)

The combination of erythromycin and rifampin (RIF) has become the standard treatment of foals with R. equi pneumonia in the 1980s and has dramatically reduced foal mortality since its introduction (Hillidge 1987, Sweeney et al. 1987). In recent years, clarithromycin or azithromycin (AZM), both newer generation macrolides, have replaced erythromycin in the combination with rifampin (Giguère et al. 2004). Compared to erythromycin, AZM has an increased stability at an acid pH, a longer half-life, and the ability to achieve high tissue concentrations by direct uptake and delivery via phagocytes (Lode et al. 1996). Clinical microbiology studies have shown that AZM has a broad spectrum of action against gram-positive and gram-negative aerobes and anaerobes because of its ability to inhibit protein synthesis at the 50s ribosome level (Davis et al. 2002). It is a good option in combination with macrolide antibiotics, as it acts synergistically with RIF (Prescott and Sweeney 1985, Weinstock and Brown 2002, Giguère et al. 2012). The dosage of RIF is given in a wide range in the text books: from 5-10 mg/kg once or twice daily (Dowling 2004, Knottenbelt and Malalana 2015, Davis 2018). A recent pharmacokinetic study has shown that a RIF dosage of 10 mg/kg per os once per day achieves concentrations above the minimum inhibitory concentration required to

inhibit the growth of 90% of all organisms (MIC90) for R. equi in the pulmonary compartments (Epithelial Lining fluid (ELF), Bronchoalveolar Lavage Cells (BALC)) (Berlin et al. 2017). The antimicrobial effect of rifampin against R. equi acts bacteriostatic, time-dependent and is additionally characterized by a long post-antibiotic effect (Giguère et al. 2012).

Tulathromycin (TUL), a novel long-acting macrolide antibiotic of the triamilide group was introduced for treatment of bacterial respiratory infections of cattle and swine in 2004 (Benchaoui et al. 2004, Traeder and Grothues 2004). A single injection provides seven days of therapeutic concentrations in lung tissues of these species. After intramuscular administration, TUL is rapidly and nearly completely absorbed from the injection site. It is widely distributed and accumulates in lung tissue, but the plasma concentrations and the lung tissue concentrations are far below the MIC for R. equi, which brings into question the value of TUL as an alternative to treat pneumonia caused by R. equi in foals (Villarino et al. 2013). But as shown by in vitro studies, TUL accumulates also in leukocytes and macrophages to concentrations several times above the serum levels. Its elimination is extremely slow with half-lives of 4–6 days (Benchaoui et al. 2004, Gáler et al. 2004, Nowakowski et al. 2004). Results of a recent study reconfirmed that TUL is as effective as standard treatment with RIF/AZM (Rutenberg et al. 2017). The only observed side effect in a recent study were 12 minor ($< 5 \, \text{cm}$) swellings developed at the injection site after a total of 279 injections of TUL (Venner et al. 2007). So it is recommended to change injection side. Further advantages of TUL are the lower price, the small injection volume and the convenient frequency of administration (only a single injection per week).

The objectives of the present study are to evaluate the comparative efficacy and survival rate of foals with pneumonia and treated with RIF/AZM versus RIF/TUL.

Materials and methods

Study population

The study was a retrospective analysis of data from a German breeding farm during 2012 to 2016. The farm has a history of endemic foal pneumonia attributable to R. equi. This pathogen was isolated from tracheobronchial aspirates of 39% (17/44) to 54% (118/217) of foals with ultrasonographic evidence of pneumonia in multiple studies performed at the farm (Venner et al. 2007a, Venner et al. 2007b, Kilian 2008, Lämmer 2010, Hagist 2016). In addition, postmortem examination of 24 foals from the same farm confirmed the presence of R. equi in lung tissue of all foals with ultrasonographic lesions (Weimar 2006).

Study design and criteria for inclusion in the treatment study

Foals 21 days of age or older with an abscess score ≥ 1 cm (definition explained below) were enrolled for participation in the study. All foals born in the seasons 2012 to 2016 (i.e. five foaling seasons) meeting criteria for inclusion were listed in a spreadsheet, and 330 foals affected with pneumonia each year were randomly selected by an online research random-

izer¹. Foals with incorrect or missing data and those that were included in previous treatment studies were excluded from the analysis, resulting in a total enrolment of 1544 foals with pneumonia.

From the age of two weeks to weaning, each foal was subjected to a physical examination and a white blood cell (WBC) count weekly. Findings of physical examination were recorded, given a score, and added to a clinical score adapted for foals with respiratory disorders (*Venner* et al. 2009). The clinical score is based on respiratory rate, tachypnoea, presence and type of nasal discharge (none, serous, mucus, purulent), mandibular lymph nodes (normal or enlarged), dyspnoea (absent or present) and auscultation of the lungs and trachea (normal versus abnormal) as described previously (*McCracken* and Slovis 2009, *Venner* et al. 2009).

Foals older than three weeks with signs of respiratory disease (cough, nasal discharge, abnormal lung sounds, dyspnoea, temperature > 39.5 °C) or with a WBC count > 13 G/L were subjected to thoracic ultrasonography using a portable unit² with a 7.5 MHz linear transducer. Pulmonary abscesses were defined as focal hypoechoic areas of consolidation of the lung tissue with a diameter larger than 1.0 cm. Furthermore, the number of pulmonary abscesses noted during an examination was documented in a lung sheet. The diameters of all consolidations detected in a foal were added to a so-called "abscess score" indicated in cm, which represented the extent of pulmonary damage. For lesions that appeared asymmetrical, the diameter was measured using the average of the largest and smallest diameter. Thoracic ultrasound examination was performed on each foal with clinical signs of respiratory disease once weekly. Furthermore, foals, with pulmonary lesions smaller than the treatment protocol requirements, were evaluated sonographically twice weekly, and closely monitored but not treated with antibiotics. Additionally, all treated foals were monitored daily for adverse reactions, such as diarrhoea, lameness, colic, or swelling at the injection site, if injections were given. A treatment duration of six weeks was set for all foals, for which treatment was deemed necessary. All foals with worsening pneumonia during the assigned treatment, either RIF/AZM or RIF/TUL, were switched to the opposite therapy. All treatments were given by different individuals not involved in monitoring and without influence on treatment assignment.

The recorded variables are summarized in Table 1.

Data analysis

Data were screened for aberrant values using range and logic checks. Foals with data errors that could not be corrected were excluded from the analysis. The distributions of continuous variables were graphically assessed using histograms and normal probability plots. Univariate comparisons of categorical variables were performed using Pearson's χ^2 test, or Fisher's exact test if expected counts were less than five. The Mann-Whitney test was used to compare treatment groups with respect to continuous variables, and the Kaplan-Meier method with the log rank test was used for univariate comparisons of survival functions.

Table 1Descriptive evaluation criteria for a random subset of
foals (n = 1,544) diagnosed with pneumonia between 2012 and
2016 on a single German breeding farm.Deskriptive Aus-
Deskriptive Aus-
wertungskriterien der randomisiert ausgewählten Fohlen (n = 1544)
mit diagnostizierter Pneumonie zwischen 2012 und 2016 auf einem
deutschen Gestijt.

Recorded var	riables
General data	Sex Date of birth
Health history	IgG level post colostrum
	Plasma transfusion and amount
	Died of R. equi infection (yes/no)
	If death from R. equi: pneumonia or extrapulmonary
	Cause of death (if not R. equi)
	Highest recorded clinical score + date + age
	Signs of pneumonia (yes/no)
Clinical	Which signs (cough, tachypnoea, fever ${>}39.5^\circ\text{C},$ nasal discharge)
parameter	Dyspnoea (yes/no) + signs of dyspnoea, tachypnoea
	Clinical score onset of therapy
	Clinical score during first/second recurrence
	Transtracheal wash culture results
Laboratory	Highest recorded WBC count + date + age
parameter	WBC count onset of therapy
	WBC first/second recurrence
	Initial ultrasound rationale
	Highest abscess score + date + age
	Highest number of abscesses
	Abscess score at the beginning of therapy
Ultrasound	Number of abscesses at the beginning of therapy
parameters	Follow-up ultrasound rationale (e.g. WBC, weekly check-up,) clinic) + date
	Abscess score during the recurrence + number of abscesses
	Duration of abscesses and no treatment
	Treated for pneumonia (yes/no)
	Treatment rationale (e.g. ultrasound score, clinic, WBC)
	Date of first lesion + age
	Date of initiation of therapy + age
	First drug (s) used
	Duration of first therapy (days)
Treatment data	Switch (yes/no) – different drugs needed
dala	Second drug (s) used
	a.) Date second therapy stopped
	b.) Duration of second therapy
	Third drug (a+b)
	Fourth drug (a+b)
	Total duration of therapy

Multivariable survival analyses were performed using the Cox proportional hazards model. Variables considered for inclusion in the multivariable analyses were year, sex, age, abscess score, clinical score, and WBC count. A manual backward elimination procedure was used for model selection, beginning with a maximum model that contained main effects for all variables that had P < 0.20 in the univariate analysis. Non significant predictors were eliminated from the maximum model in a stepwise manner until only variables having P < 0.05 remained. Treatment was included as a predictor in all models regardless of its significance, and all two-way interactions with the treatment variable were evaluated. If the removal of any variable resulted in greater than a 10% change to the estimated hazard ratio for treatment, that variable was considered an important confounder, and it was retained in the final model. Continuous variables were assessed as both continuous and categorical predictors, with Akaike's information criterion being used to determine which form of the predictor provided the best fit. The proportional hazards assumption was assessed by evaluating the slope of the scaled Schoenfeld residuals, and by evaluating the significance of an interaction between each of the covariates and a function of time. Variables that failed to meet the proportional hazards assumption were included in the analysis by allowing them to interact with a function of time in an extended Cox model, or they were included in the analysis as a stratification variable. Data were further screened for outliers and influential observations using index plots of the deviance residuals and delta-beta values.

All statistical tests assumed a two-sided alternative hypothesis, and values of P < 0.05 were considered significant. Analyses were performed using commercially available statistical software (Stata version 15.1, StataCorp LLC, College Station, TX).

Results

Records of 1,544 foals were included in the study, with the number of foals for each of the five years ranging between 297 and 319. Seven hundred sixty-one (49.3%) foals were fillies, and 783 (50.7%) were colts. Seven hundred eightysix (50.9%) of the foals were treated for R. equi pneumonia. Of the foals that were treated, 353 (44.9%) received rifampin and azithromycin (RIF/AZM) as their first treatment, 406 (51.7%) received rifampin and tulathromycin (RIF/TUL), and 27 (3.4%) received a different drug treatment combination. The proportions of foals receiving RIF/AZM or RIF/TUL varied by year (P < 0.001) but not by sex (P = 0.52; Table 2). At the beginning of treatment, foals receiving RIF/AZM did not differ from foals receiving RIF/TUL with respect to age (P = 0.32), clinical score (P = 0.31), or WBC count (P = 0.20), but foals receiving RIF/AZM did have a higher median abscess score (P < 0.001; Table 3).

Treatment failure was defined as a worsening clinical condition that required switching to a different drug treatment protocol, or death due to R. equi pneumonia. In a univariate comparison of the two treatments, there was a higher incidence of treatment failure in foals treated with RIF/AZM (32/353, 9.1%) than in foals treated with RIF/TUL (19/406, 4.7%; Pearson's χ^2 , P = 0.016). In a Kaplan-Meier analysis, there was also a significant difference between the two treatments with respect to their survival functions (Log-rank test, P = 0.045; Figure 1). The incidence of mortality due to R. equi pneumonia was higher in foals treated with RIF/AZM (8/353, 2.3%) than in those treated with RIF/TUL (0/406, 0.0%; Fisher's exact test, P = 0.002).

Results of a multivariable Cox regression analysis for treatment failure are summarized in Table 4. After adjusting for year, age of foals at the beginning of treatment, and abscess score at the beginning of treatment, the risk of treatment failure was not significantly lower for foals treated with RIF/TUL compared to those treated with RIF/AZM [Hazard Ratio (95%

Table 2Number (%) of foals treated with rifampin and azithro-
mycin (RIF/AZM) or rifampin and tulathromycin (RIF/TUL) by year and
sex. | Anzahl (%) der Fohlen behandelt mit Rifampin und Azithromy-
cin (RIF/AZM) beziehungsweise Rifampin und Tulathromycin (RIF/TUL)
sortiert nach Jahr und Geschlecht.

	RIF/AZM (n = 353)	RIF/TUL (n = 406)	† <i>P</i>
Year			< 0.001
2012	33 (18.2)	148 (81.8)	
2013	138 (63.9)	78 (36.1)	
2014	36 (30.5)	82 (69.5)	
2015	98 (56.3)	76 (43.7)	
2016	48 (68.6)	22 (31.4)	
Sex			0.519
Female	163 (47.8)	178 (52.2)	
Male	190 (45.5)	228 (54.5)	

+P-value for Pearson's $\chi 2$ test

 Table 3
 Clinical characteristics of foals treated with rifampin and azithromycin (RIF/AZM) or rifampin and tulathromycin (RIF/TUL) at the beginning of treatment.
 Klinische Parameter der Fohlen behandelt mit Rifampin und Azithromycin (RIF/AZM) beziehungsweise Rifampin und Tulathromycin (RIF/TUL) zu Beginn der Behandlung.

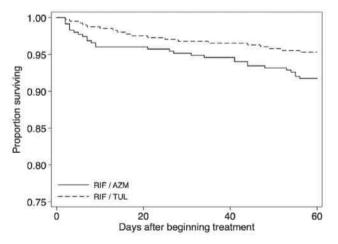
	RIF/AZM (n = 353)	RIF/TUL (n = 406)	†P
Age (days)			0.320
Mean (SD)	106 (38)	105 (30)	
Median (Min, Max)	109 (26, 231)	106 (33, 205)	
Abscess score (cm)			< 0.001
Mean (SD)	15.8 (9.8)	11.7 (4.4)	
Median (Min, Max)	13.5 (0, 64)	11.0 (0, 30)	
Clinical score			0.313
Mean (SD)	2.2 (1.1)	2.1 (1.1)	
Median (Min, Max)	2 (0, 6)	2 (0, 6)	
WBC count $\times 10^3$ (cells	0.198		
Mean (SD)	15.0 (4.7)	15.0 (4.4)	
Median (Min, Max)	14.3 (4.2, 34.7)	14.8 (3.8, 23.4)	

†P-value for the Mann-Whitney test

CI) = 0.75 (0.38, 1.5)]. Age of foals was negatively associated with treatment failure, with the risk of failure decreasing by 30% for every one-month increase in age at the beginning of treatment [HR (95% CI) = 0.70 (0.54, 0.91)]. Abscess score was positively associated with treatment failure, with the risk of failure increasing by 4% for every one cm increase in score [HR (95% CI) = 1.04 (1.01, 1.07)]. Year was also associated with the risk of treatment failure but it did not meet the proportional hazards assumption, so the model was adjusted for the effects of year by stratification.

If foals required a second round of treatment for pneumonia after the end of the initial treatment period, they were considered to have had a recurrence. In a univariate comparison of the two treatments, there was a higher incidence of recurrence in foals treated with RIF/AZM (59/349, 16.9%) than in foals that were treated with RIF/TUL (28/399, 7.0%; Pearson's χ^2 , P < 0.001). In a Kaplan-Meier analysis, there was also a significant difference between the two treatments with respect to their survival functions (Log rank test, P < 0.001; Figure 2).

Results of a multivariable Cox regression analysis for recurrence of pneumonia are summarized in Table 5. After adjusting for year and age of the foals at the end of their initial treatment period, the risk of recurrence was 54% lower in foals treated with RIF/TUL compared to foals that were treated with RIF/AZM [HR (95% CI) = 0.46 (0.28, 0.76)]. Age of foals was negatively associated with recurrence, with the risk of recurrence decreasing by 65% for every one-month increase in age. The proportional hazards assumption was not met for year, so, the model was adjusted for the effects of year by stratification.



Kaplan-Meier graph illustrating time from the beainning Fig. 1 of treatment until treatment failure in foals treated with rifampin and azithromycin (RIF/AZM) or rifampin and tulathromycin (RIF/TUL). Treatment failure was defined as a worsening clinical condition that required switching to a different drug treatment protocol, or death due to R. equi pneumonia. (X: Days after treatment onset, Y: Survival Kaplan-Meier-Kurve zur Veranschaulichung der Zeit von rate) Therapiebeginn bis Therapieversagen bei Fohlen behandelt mit Rifampin und Azithromycin (RIF/AZM) beziehungsweise Rifampin und Tulathromycin (RIF/TUL). Therapieversagen wurde definiert als Verschlechterung des klinischen Zustandes, der Therapieumstellung auf ein anderes Behandlungsprotokoll erforderte, oder den Tod durch R. equi Pneumonie. (X: Tage nach Behandlungsbeginn, Y: Überlebensrate)

Discussion

As results of previous studies have shown, RIF alone is insufficient as an R. equi therapy thus it should not be used alone due to emerging resistant strains (*Prescott* and *Nicholson* 1984, *Nordmann* and *Ronco* 1992, *Giguère* et al. 2010, *Berghaus* et al. 2013). The combination of RIF with a macrolide antibiotic has proved to be very effective in in vitro studies as well as in foals with natural infection with R. equi (*Sweeney* 1987, *Giguère* et al 2012. *Hildebrandt* 2015, *Rutenberg* 2017). The efficacy of several macrolides has been evaluated in the treat-

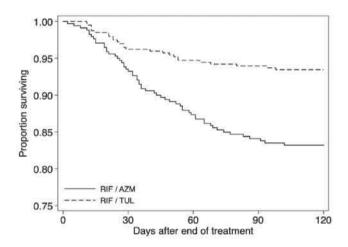


Fig. 2 Kaplan-Meier graph illustrating time from the end of the initial treatment period until pneumonia recurrence in foals treated with rifampin and azithromycin (RIF/AZM) or rifampin and tulathromycin (RIF/TUL). (X: Days after treatment onset, Y: Survival rate) | Kaplan-Meier-Kurve zur Veranschaulichung der Zeit vom Ende der initialen Behandlungsperiode bis zu einem Rückfall der mit Rifampin und Azithromycin (RIF/AZM) beziehungsweise Rifampin und Tulathromycin (RIF/TUL) behandelten Fohlen. (X: Tage nach Behandlungsbeginn, Y: Überlebensrate)

Table 4Summary of a stratified* multivariable Cox regression mod-
el for time from the beginning of treatment until treatment failure in foals
(n = 754) treated with rifampin and azithromycin (RIF/AZM) or rifampin
and tulathromycin (RIF/TUL). Treatment failure was defined as a worsen-
ing clinical condition that required switching to a different drug treatment
protocol, or death due to R. equi pneumonia. | Zusammenfassung der
stratifizierten* multivariablen Cox-Regression der Zeit von Therapiebeginn
bis zum Therapieversagen der Fohlen (n = 754) behandelt mit Rifampin
und Azithromycin (RIF/AZM) beziehungsweise Rifampin und Tulathromycin
(RIF/TUL). Therapieversagen wurde definiert als Verschlechterung des kli-
nischen Zustandes, der Therapieumstellung auf ein anderes Behandlungs-
protokoll erforderte oder den Tod durch R. equi Pneumonie.

Variable	Coefficient (SE)	Hazard Ratio (95% CI)	†P
Treatment RIF/AZM RIF/TUL	Reference 0.287 (0.343)	Reference 0.75 (0.38, 1.5)	0.401
Age (months) at beginning of treatment	-0.355 (0.135)	0.70 (0.54, 0.91)	0.009
Abscess score (cm) at begin- ning of treatment	0.040 (0.015)	1.04 (1.01, 1.07)	0.007

*Stratified by year, †Wald P-value

Table 5Summary of a stratified* multivariable Cox regressionmodel for time from the end of the initial treatment period until pneu-
monia recurrence in foals (n = 737) treated with rifampin and azi-
thromycin (RIF/AZM) or rifampin and tulathromycin (RIF/TUL).Zusammenfassung der stratifizierten* multivariablen Cox-Regression
der Zeit vom Ende der initialen Behandlungsperiode bis zu einem
Rückfall der mit Rifampin und Azithromycin (RIF/AZM) beziehungs-
weise Rifampin und Tulathromycin (RIF/TUL) behandelten Fohlen
(n = 737).

Variable	Coefficient (SE)	Hazard Ratio (95% CI)	†P
Treatment			0.002
RIF/AZM RIF/TUL	Reference –0.784 (0.257)	Reference 0.46 (0.28, 0.76)	
Age (months) at end of initial treatment period	–1.057 (0.106)	0.35 (0.28, 0.43)	< 0.001

*Stratified by year, †Wald P-value

ment of R. equi pneumonia in foals in recent years. RIF/ TUL appears to be an appropriate therapy of foals with R. equi pneumonia due to several reasons. From the antibiotics used in this study, TUL is the only drug licensed for veterinary use. In terms of the drug cascade, it should be the first choice of the veterinarian before using drugs only licensed for human medicine. Additionally, the therapy interval with injections every seven days is simple to manage, and is well tolerated by the foals (Venner et al. 2007). Minor side effects observed are rare and most of the time self-limiting. Furthermore TUL itself is less expensive than azithromycin and, because it is given parenterally, it is less likely to contaminate the environment than orally administered AZM. The data analyzed here shows that the number of foals treated with RIF/TUL was higher during some years compared to other periods, as seen in Table 3. This is due to the fact that the treatment protocols changed over the years due to results of several treatment studies or individual veterinary decisions. That makes it difficult to directly compare the efficacy of RIF/TUL and RIF/AZM even if statistical adjustments for abscess score, clinical score and age were made in the present study.

Nevertheless, the present analysis of a large data set suggests that for the treatment of mild to moderate R. equi pneumonia, the combination of RIF/TUL seems adequate and leaves the veterinarian with the powerful combination of RIF/AZM in reserve in the rare cases when the pneumonia worsens. Finally, the option of treating foals with different severity levels of pneumonia with an alternative antibiotic combination has a benefit for the foal and the owner.

In conclusion, rifampin/tulathromycin is an effective treatment for foals with mild to moderate pneumonia.

Footnotes

² Esaote Tringa Linear, Milano Italy.

Conflict of interest

The authors declare that there is no conflict of interest.

References

- Burton A. J., Giguère S., Sturgill T. S., Berghaus L. J., Slovis N. M., Whitman J. L., Levering C., Kuskie K. R., Cohen N. D. (2013) Macrolide- and Rifampin-Resistant Rhodococcus equi on a Horse Breeding Farm, Kentucky, USA. Emerg. Infect. Dis. 19, 282; DOI 10.3201/eid1902.121210
- Benchaoui H. A., Nowakowski M., Sherington J., Rowan T. G., Sunderland S. J. (2004) Pharmacokinetics and lung tissue concentrations of tulathromycin in swine. J. Vet. Pharmacol. Ther. 27, 203– 210; DOI 10.1111/j.1365-2885.2004.00586.x
- Berghaus L. J., Giguère S., Guldbech K. (2013) Mutant prevention concentration and mutant selection window for 10 antimicrobial agents against Rhodococcus equi. Vet. Microbiol. 166, 670–675; DOI 10.1016/j.vetmic.2013.07.006
- Berlin S., Spieckermann L., Oswald S., Berlin S., Keiser M., Lumpe S., Ullrich A., Grube M., Hasan M., Venner M., Siegmund W. (2016) Pharmacokinetics and Pulmonary Distribution of Clarithromycin and Rifampicin after Concomitant and Consecutive Administration in Foals. Mol. Pharm. 13, 1089–1099; DOI 10.1021/acs.molpharmaceut.5b00907
- Berlin S., Kirschbaum A., Spieckermann L., Oswald S., Keiser M., Grube M., Venner M., Siegmund W. (2017) Pharmacological indices and pulmonary distribution of rifampicin after repeated oral administration in healthy foals. Equine Vet. J. 49, 618–623; DOI 10.1111/evj.12662
- Cohen N. D. (2014) Rhodococcus equi Foal Pneumonia. Vet. Clin. North Am. Equine Pract. 30, 609–622; DOI 10.1016/j.cveq.2014.08.010
- Davis J. L., Gardner S. Y., Jones S. L., Schwabenton B. A., Papich M. G. (2002) Pharmacokinetics of azithromycin in foals after i.v. and oral dose and disposition into phagocytes. J. Vet. Pharmacol. Ther. 25, 99–104; DOI 10.1046/j.1365-2885.2002.00387.x
- Davis E. (2018) Disorders of the Respiratory System. Equine Internal Medicine. Reed S. M., Bayly W. M., Sellon D. C. Elsevier, Amsterdam, Fourth Edition, 313–386; DOI 10.1016/B978-0-323-44329-6.00008-5
- Dowling P. M. (2004) Antimicrobial therapy. Equine Clinical Pharmacology. Bertone J. J., Horspool L. J. I. Saunders Ltd., Philadelphia. First Edition, 13–47; DOI 10.1016/B978-0-7020-2484-9.50006-7
- Gáler D., Hessong S., Beato B. Risk J., Inskeep P., Weerasinghe C., Schneider R. P., Langer C., LaPerle J., Renouf D., Bessire A., Español E., Rafka R., Ragan C., Boettner W., Murphy T., Keller D., Benchaoui H., Nowakowski M. A. (2004) An Analytical Method for the Analysis of Tulathromycin, an Equilibrating Triamilide, in Bovine and Porcine Plasma and Lung. J. Agr. Food Chem. 52, 2179–2191; DOI 10.1021/jf0351624
- Gaynor M., Mankin A. S. (2003) Macrolide antibiotics: binding site, mechanism of action, resistance. Curr. Top. Med. Chem. 3, 949– 961; DOI 10.2174/1568026033452159
- Giguère S., Berghaus L. J., Lee E. A. (2015) Activity of 10 antimicrobial agents against intracellular Rhodococcus equi. Vet. Microbiol. 178, 275–278; DOI 10.1016/j.vetmic.2015.05.019
- Giguère S., Lee E., Williams E., Cohen N. D., Chaffin M. K., Halbert N., Martens R. J., Franklin R. P., Clark C. C., Slovis N. M. (2010) Determination of the prevalence of antimicrobial resistance to macrolide antimicrobials or rifampin in Rhodococcus equi isolates and treatment outcome in foals infected with antimicrobial-resistant isolates of R equi. J. Am. Vet. Med. Assoc. 237, 74–81; DOI 10.2460/javma.237.1.74
- Giguère S., Lee E. A., Guldbech K. M., Berghaus L. J. (2012) In vitro synergy, pharmacodynamics, and postantibiotic effect of 11 antimicrobial agents against Rhodococcus equi. Vet. microbiol. 160, 207–213; DOI 10.1016/j.vetmic.2012.05.031

¹ https://www.randomizer.org

- Giguère S., Jacks S., Roberts G. D., Hernandez J., Long M. T., Ellis C. (2004) Retrospective Comparison of Azithromycin, Clarithromycin, and Erythromycin for the Treatment of Foals with Rhodococcus equi Pneumonia. J. Vet. Intern. Med. 18, 568–573; DOI 10.1111/j.1939-1676.2004.tb02587.x
- Hagist C. (2016) Genotypisierung von Rhodococcus equi Stämmen aus Deutschland, isoliert bei Fohlen und anderen Tierarten. Diss. Med. Vet. Hannover, 134
- Hildebrand F, Venner M., Giguère S. (2015) Efficacy of Gamithromycin for the Treatment of Foals with Mild to Moderate Bronchopneumonia. J. Vet. Intern. Med 29, 333–338; DOI 10.1111/ jvim.12504
- Hillidge C. J. (1987) Use of erythromycin-rifampin combination in treatment of Rhodococcus equi pneumonia. Vet. Microbiol. 14, 337–342; DOI 10.1016/0378-1135(87)90121-0
- Hoffman A. M., Viel L., Prescott J. F., Rosendal S., Thorsen J. (1993) Association of microbiologic flora with clinical, endoscopic, and pulmonary cytologic findings in foals with distal respiratory tract infection. Am. J. Vet. Res. 54, 1615–1622
- *Kilian K.* (2008) Vergleichende Untersuchungen zum Nachweis von Rhodococcus equi in der Atemluft, im Tracheobronchialsekret und im Kot von Fohlen. Diss. Med. Vet. Hannover, 128.
- Knottenbelt D. C., Malalana F. (2015) Part 3 Index of Drugs Used in Equine Medicine. Saunders Equine Formulary. Knottenbelt D. C., Malalana F., W. B. Saunders. Second Edition, 60–257; DOI 10.1016/B978-0-7020-5109-8.00003-1
- Lode H., Borner K., Koeppe P., Schaberg T. (1996) Azithromycin-review of key chemical, pharmacokinetic and microbiological features. J. Antimicrob. Chemother. 37, 1–8; DOI 10.1093/ jac/37
- Lämmer M. (2010) Nachweis von Rhodococcus equi in Kot und Trachealsekret bei Fohlen: Vergleichende Untersuchung zwischen gesunden Fohlen und Fohlen mit Lungenabszessen. Diss. Med. Vet. Hannover, 116
- McCracken J. L., Slovis N. M. (2009) Use of thoracic ultrasound for the prevention of Rhodococcus equi pneumonia on endemic farms. Proc. Am. Assoc. Equine Pract. 55, 38–44.
- Muscatello G. (2012) Rhodococcus equi pneumonia in the foal Part 1: Pathogenesis and epidemiology. Vet. J. 192, 20–26; DOI 10.1016/j.tvjl.2011.08.014
- Muscatello G. (2012) Rhodococcus equi pneumonia in the foal Part 2: Diagnostics, treatment and disease management. Vet. J. 192, 27–33; DOI 10.1016/j.tvjl.2011.08.009
- Nordmann P, Ronco E. (1992) In-vitro antimicrobial susceptibility of Rhodococcus equi. J. Antimicrob. Chemother 29, 383–393; DOI 10.1093/jac/29.4.383
- Nowakowski M. A., Inskeep P. B., Risk J. E., Skogerboe T. L., Benchaoui, H. A., Meinert T. R., Sherington J., Sunderland S. J. (2004) Pharmacokinetics and lung tissue concentrations of tulathromycin, a new triamilide antibiotic, in cattle. Vet. Therapeut. 5, 60–74.

- Peters J., Block W., Oswald S. Freyer J., Grube M., Kroemer H. K., Lämmer M., Lutjohann D., Venner M., Siegmund W. (2011) Oral absorption of clarithromycin is nearly abolished by chronic comedication of rifampicin in foals. Drug. Metab. Dispos. 39, 1643–1649; DOI 10.1124/dmd.111.039206
- Prescott J. F., Nicholson V. M. (1984) The effects of combinations of selected antibiotics on the growth of Corynebacterium equi. J. Vet. Pharmacol. Ther. 7, 61–64; DOI 10.1111/j.1365-2885.1984.tb00880.x
- Prescott J. F., Sweeney C. R. (1985) Treatment of Corynebacterium equi pneumonia of foals: a review. J. Am. Vet. Med. A. 187, 725–728.
- Rutenberg D., Venner M., Giguère S. (2017) Efficacy of Tulathromycin for the Treatment of Foals with Mild to Moderate Bronchopneumonia. J. Vet. Intern. Med 31, 901–906; DOI 10.1111/jvim.14717
- Sweeney C. R., Sweeney R. W., Divers T. J. (1987) Rhodococcus equi pneumonia in 48 foals: response to antimicrobial therapy. Vet. Microbiol. 14, 329–336; DOI 10.1016/0378-1135(87)90120-9
- Traeder W., Grothues M. (2004) Pharmacological characteristics and efficacy of Tulathromycin, the first representative of the Triamilide antibiotics. Tierärztliche Umschau 59 (2),102–113
- Vázquez-Boland J. A., Letek M., Valero-Rello A., González P., Scortti M., Fogarty U. (2010) Rhodococcus equi and Its Pathogenic Mechanisms. Biology of Rhodococcus. Microbiology Monographs. Alvarez H. Volume 16, 331–359
- Venner M., Heyers P., Strutzberg-Minder K., Lorenz, N., Verspohl J., Klug E. (2007) Detection of rhodococcus equi by microbiological culture and by polymerase chain reaction in samples of tracheobronchial secretions of foals. Berl. Münch. Tierärztl. Wschr. 120, 126–133; DOI 10.2376/0005-9366-120-126
- Venner M. Kerth R., Klug E. (2007a) Evaluation of tulathromycin in the treatment of pulmonary abscesses in foals. Vet. J. 174, 418– 421; DOI 10.1016/j.tvjl.2006.08.016
- Venner M., Meyer-Hamme B., Verspohl J., Hatori, F., Shimizu N., Sasaki Y., Kakuda T., Tsubaki S., Takai S. (2007b) Genotypic characterization of VapA positive Rhodococcus equi in foals with pulmonary affection and their soil environment on a warmblood horse breeding farm in Germany. Res. Vet. Sci. 83, 311–317; DOI 10.1016/j.rvsc.2007.01.009
- Venner M., Reinhold B., Beyerbach M., Feige K. (2009) Efficacy of azithromycin in preventing pulmonary abscesses in foals. Vet. J. 179, 301–303; DOI 10.1016/j.tvjl.2007.10.002
- Villarino N., Brown S. A., Martín-Jiménez T. (2013) The role of the macrolide tulathromycin in veterinary medicine. Vet. J. 198, 352– 357; DOI 10.1111/jvp.12010
- Weimar B. M. (2006) Lungenabszesse bei Fohlen: Klinische, sonographische, endoskopischen, pathomorphologische und mikrobiologische Befunde. Diss. Med. Vet. Hannover, 157
- Weese J. S., Giguère S., Guardabassi L., Morley P. S., Papich M., Ricciuto D. R., Sykes J. E. (2015) ACVIM consensus statement on therapeutic antimicrobial use in animals and antimicrobial resistance. J. Vet. Intern. Med 29, 487–498; DOI 10.1111/ jvim.12562
- Weinstock D. M., Brown A. E. (2002) Rhodococcus equi: an emerging pathogen. Clin. Infect. Dis. 34, 1379–1385; DOI 10.1086/340259